

# Transforming Growth Factor- $\beta$ in Normal Nociceptive Processing and Pathological Pain Models

Aquilino Lantero · Mónica Tramullas · Alvaro Díaz ·  
María A. Hurlé

Received: 17 September 2011 / Accepted: 9 November 2011 / Published online: 29 November 2011  
© Springer Science+Business Media, LLC 2011

**Summary** The transforming growth factor- $\beta$  (TGF- $\beta$ ) superfamily is a multifunctional, contextually acting family of cytokines that participate in the regulation of development, disease and tissue repair in the nervous system. The TGF- $\beta$  family is composed of several members, including TGF- $\beta$ s, bone morphogenetic proteins (BMPs) and activins. In this review, we discuss recent findings that suggest TGF- $\beta$  function as important pleiotropic modulators of nociceptive processing both physiologically and under pathological painful conditions. The strategy of increasing TGF- $\beta$  signaling by deleting “BMP and activin membrane-bound inhibitor” (BAMBI), a TGF- $\beta$  pseudoreceptor, has demonstrated the inhibitory role of TGF- $\beta$  signaling pathways in normal nociception and in inflammatory and neuropathic pain models. In particular, strong evidence suggests that TGF- $\beta$ 1 is a relevant mediator of nociception and has protective effects against the development of chronic neuropathic pain by

inhibiting the neuroimmune responses of neurons and glia and promoting the expression of endogenous opioids within the spinal cord. In the peripheral nervous system, activins and BMPs function as target-derived differentiation factors that determine and maintain the phenotypic identity and circuit assembly of peptidergic nociceptors. In this context, activin is involved in the complex events of neuroinflammation that modulate the expression of pain during wound healing. These findings have provided new insights into the physiopathology of nociception. Moreover, specific members of the TGF- $\beta$  family and their signaling effectors and modulator molecules may be promising molecular targets for novel therapeutic agents for pain management.

**Keywords** TGF- $\beta$  · Activin · BMP · BAMBI · Nociception · Neuropathic pain · Pain

A. Lantero · M. Tramullas · A. Díaz · M. A. Hurlé  
Departamento de Fisiología y Farmacología,  
Facultad de Medicina, Universidad de Cantabria,  
39011 Santander, Spain

A. Lantero · M. Tramullas · M. A. Hurlé (✉)  
Instituto de Formación e Investigación  
Marqués de Valdecilla (IFIMAV),  
39011 Santander, Spain  
e-mail: hurlem@unican.es

A. Díaz  
Instituto de Biomedicina y Biotecnología  
de Cantabria IBBTEC (UC-CSIC-IDICAN),  
Santander, Spain

A. Díaz  
Centro de Investigación Biomédica en Red de  
Salud Mental (CIBERSAM), Instituto de Salud Carlos III,  
Madrid, Spain

## Introduction

Pain is an unpleasant but indispensable sensation that warns the body to protect itself from severe damage. However, prolonged suffering from pain becomes a serious burden, and it is a major reason for which patients seek medical care and pharmacological treatment. Chronic pain is remarkably prevalent; it affects millions of people worldwide. Epidemiological surveys have revealed the staggering extent of chronic pain, its human costs and socio-economic impact, and also the paucity of effective methods for pain control [1, 2]. Poor pain control is the result of a deficit in our understanding of the mechanisms of chronic pain, which has limited our arsenal of pathogenesis-based analgesic therapies. However, there is considerable hope for the development of

new classes of analgesic drugs that target novel processes that contribute to clinically relevant pain. In this review, we highlighted recent results that identified TGF- $\beta$ s as important pleiotropic modulators of nociceptive processing in physiological and pathological pain. The evidence indicates that specific members of the TGF- $\beta$  family and their signaling effectors and modulators may be promising molecular targets for novel therapeutic agents for pain management.

### The transforming Growth Factor- $\beta$ Superfamily of Cytokines

The transforming growth factor- $\beta$  (TGF- $\beta$ ) superfamily is a multifunctional, contextually acting family of cytokines that is comprised of more than 30 proteins. In mammals, these cytokines are grouped into different subfamilies: TGF- $\beta$ s, bone morphogenetic proteins (BMPs), activins, growth and differentiation factors, anti-müllerian hormone and nodal. Members of the TGF- $\beta$  family are produced as large precursor proteins within the cell, and they elicit biological responses as extracellularly secreted homo- or heterodimers. TGF- $\beta$ s signal through serine (ser)-threonine (thr) kinase receptors of type I or activin receptor-like kinases (ALKs) and type II receptors (T $\beta$ R-II). Ligand binding induces the formation of a stable receptor heterotetrameric complex that is composed of two receptors of each type. TGF- $\beta$  family receptors are shared among ligands; only five type II receptors and seven type I receptors were described. Each member of the TGF- $\beta$  superfamily binds to a characteristic combination of type I and type II receptors (Table 1). Type II receptors have intrinsic ser/thr protein kinase activity and transphosphorylate type I receptors at their GS domains (glycine- and serine-rich sequences), which stimulate their protein kinase activity; this phosphorylation is necessary and sufficient for TGF- $\beta$  signaling. A type III receptor (T $\beta$ R-III), also known as betaglycan, has also been described. However, type III receptors lack signaling domain and appear to act as co-receptors that enhance the binding of TGF- $\beta$ s and BMPs to type I or type II receptors [3].

### Canonical TGF- $\beta$ Signaling

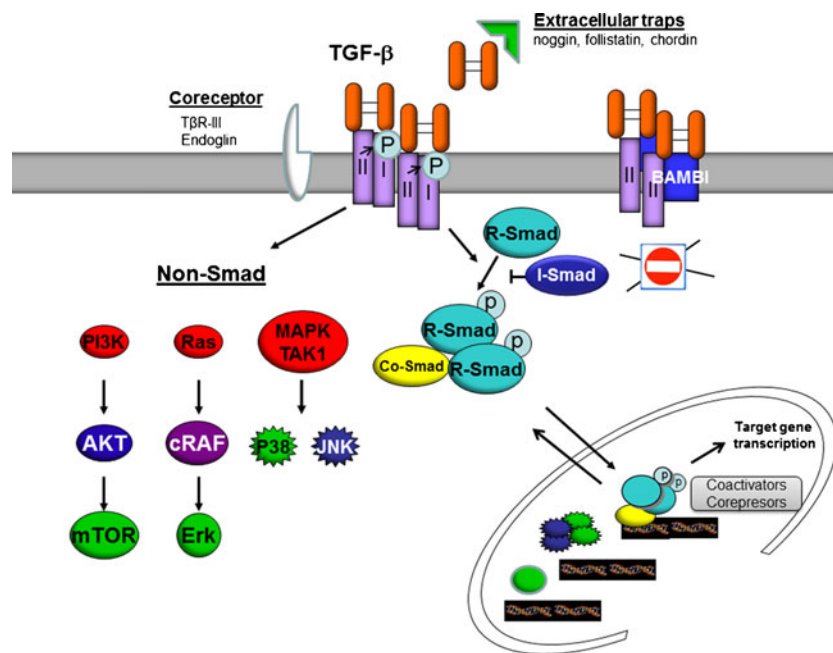
The activation of type I receptor kinases leads to the downstream propagation of the signal through the phosphorylation of intracellular receptor-activated Smad (R-Smad) proteins (Table 1, Fig. 1). Phosphorylated R-Smads interact with co-Smad4, which is a common co-factor of all TGF- $\beta$  activin and BMP signaling pathways, to form Smad complexes that translocate to the nucleus and regulate gene transcription. The interaction of the type I receptor with specific R-Smad proteins is dependent on the ligand. TGF- $\beta$ s and activins signal via Smad2 and Smad3, and BMPs signal through Smad1, Smad5 and Smad8. Smad complexes bind specific DNA sequences in the regulatory regions of numerous target genes, but the recruitment of Smads to a particular promoter and the specific transcriptional response that is elicited depend on the interaction of Smads with various DNA binding co-factors, co-activators and co-repressors [4]. This cooperative interaction between Smads and their DNA-binding partners confers a large spectrum of sequence specificities to TGF- $\beta$  signaling and the potential for the integration or interaction of multiple signaling pathways within the cell [5].

### Non-canonical TGF- $\beta$ Signaling

TGF- $\beta$  also alters cell behavior through the activation of Smad-independent pathways (Fig. 1). The mechanism that couples TGF- $\beta$  to non-canonical effector systems and the biological consequences of this coupling remain poorly characterized. Several non-Smad signaling pathways with links to the TGF- $\beta$  receptor complex are partially understood, including MAP kinases [TGF- $\beta$ -activated kinase 1 (TAK1), Erk, p38 MAPK and c-Jun N-terminal kinase (JNK)], calcium-dependent phosphatase calcineurin-NFATc, growth and survival kinases PI3K/AKT/mTOR and small GTP-binding proteins Ras, RhoA, RhoB, Rac1 and Cdc42. The nuclear signals that are transmitted by non-Smad proteins can regulate transcription independently or synergize with the Smad proteins. Non-Smad proteins modulate the activity and signaling of Smad proteins. Furthermore, there is extensive crosstalk between Smads and

**Table 1** Ligand–receptor–Smad relationships in the TGF- $\beta$  family

Ligand	Type II receptors	Type I receptors	R-Smad	Co-Smad	I-Smads
TGF- $\beta$ s	T $\beta$ R-II	ALK5/T $\beta$ R-I ALK1	Smad2 Smad3 Smad1 Smad5 Smad8	Smad4	Smad7
Activins	ActRIIA ActRIIB	ALK4/ActR1B ALK7/ActR1C	Smad2 Smad3	Smad4	Smad7
BMPs	BMPR-II ActRIIA ActRIIB	ALK3/BMPR-IA ALK6/BMPR-IB ALK1ALK2/ActR1A	Smad1 Smad5 Smad8	Smad4	Smad6 Smad7



**Fig. 1** The general mechanism of TGF- $\beta$  receptor activation in Smad and non-Smad signaling pathways. The binding of ligands triggers heteromeric complex formation between TGF- $\beta$  type I and type II receptors. Type I receptor is transphosphorylated and activated by the type II receptor kinase. The activated type I receptors phosphorylate R-Smads. Activated R-Smads form a complex with a common Smad4. R-Smad/Smad4 complexes translocate into the nucleus, where they

regulate the transcription of target genes. The activation of R-Smads is inhibited by Smad6 or Smad7. TGF- $\beta$ s also activate non-Smad signaling pathways, such as MAPK/p38/JNK and Ras/Erk-MAPK. TGF- $\beta$  pathways are regulated by molecules that bind ligands in the extracellular space. Several membrane-associated proteins modulate the reception of TGF- $\beta$  signals by the cell. BAMBI is a decoy receptor that prevents downstream signal transduction

kinase effectors with activity that is modulated by TGF- $\beta$  ligands [6].

### Regulation of TGF- $\beta$ Signaling

The cell type- and context-dependent biological responses elicited by TGF- $\beta$ s are determined by a variety of factors, including the extracellular concentration of the ligand, the presence and quantity of the complementary receptor on the target cell surface and the downstream signals that are activated [7]. The pleiotropic nature of TGF- $\beta$ s is achieved through a tight control and a fine-tune of the strength, positioning and timing of signaling. Multiple mechanisms at every level, from the extracellular space to the transcriptional activity in the nucleus, cause remarkable context-specific gains or losses in TGF- $\beta$  signaling [8, 9].

TGF- $\beta$  pathways are often regulated by molecules that bind specific ligands in the extracellular space to limit their availability, control their diffusion from the producing cells, affect their transit through tissues or block their binding to receptors (Fig. 1). These regulatory molecules include a large set of specific extracellular diffusible proteins, such as noggin, follistatin, chordin, Dan, cerberus/caronte and gremlin [9].

Several membrane-associated proteins also modulate the cellular reception of TGF- $\beta$  signals. The BMP and activin

membrane-bound inhibitor (BAMBI) is structurally similar to type I receptors, but it lacks an intracellular kinase domain. Consequently, BAMBI acts as a decoy type I receptor that negatively modulates TGF- $\beta$ /BMP/activin signaling by stably associating with type II receptors, which prevents the formation of active receptor complexes (Fig. 1) [10].

TGF- $\beta$ -bound receptor complexes internalize by endocytosis. Clathrin-mediated internalization is required for Smad activation by the receptor complex, whereas caveolin- and lipid-raft-mediated endocytosis has been associated with receptor degradation. Protein associations that regulate the selection of the routing by the receptors would define the strength and duration of the signals and responses, and the receptor turnover [7].

The inhibitory Smad6 and Smad7 (I-Smads) are structurally divergent Smads which negatively regulate signaling strength and duration (Fig. 1). I-Smads bind to type I receptors and competitively inhibit R-Smad phosphorylation and the recruitment of phosphatases and Smurf ubiquitin ligases to downregulate receptor levels and activity [8].

A broad array of cytoplasmic and nuclear Smad interaction partners regulates Smad responses by (1) modulating their recruitment to the receptor complex, (2) controlling their phosphorylation, dephosphorylation and

sumoylation, (3) sequestering Smads from active signaling participation or (4) modulating the association of the R-Smad/Smad4 complex with transcription factors, co-activators and co-repressors and, subsequently, the Smad-dependent transcription [11], among other mechanisms.

## Regulatory Roles of the TGF- $\beta$ Family in Normal Nociceptive Processing and Pathological Pain Models

### Nociception Overview

Nociception is a specialized form of sensory signaling that conveys information about potentially damaging stimuli to the central nervous system (CNS). The transduction of noxious stimuli originates at the peripheral axon terminals of high-threshold unmyelinated C or thinly myelinated A $\delta$  primary sensory neurons that innervate the target tissue. Functional and molecular nociceptor heterogeneity is associated with the specific detection of distinct pain modalities. Nociceptor cell bodies reside in the dorsal root ganglia (DRG) of spinal nerves and the sensory ganglia of cranial nerves. The primary afferent projections from nociceptors transmit the signal from the periphery to the spinal cord dorsal horn, where they form synapses with second-order neurons in laminae I and V. The ascending fibers of second-order neurons project to third-order neurons in the thalamus and brainstem, which transmit the nociceptive information to higher brain structures that interpret the sensory-discriminative, affective-emotional or aversive dimensions of pain [12].

The release of inflammatory mediators within the wounded area amplifies peripheral nociceptor transduction. This “peripheral sensitization” is a form of stimulus-evoked functional plasticity, which normally occurs during the healing process, in which tissue damage enhances the excitability of nociceptors to protect the injured area by increasing pain sensitivity [13]. This physiological “nociceptive” pain normally disappears upon healing. However, pain persists in a state of chronic neuropathic pain after nervous system injuries in a small but significant percentage of the population [14].

In chronic neuropathic pain, a shift in the balance between the excitatory and inhibitory mechanisms that modulate spinal cord excitability heightens the response of dorsal horn neurons to incoming afferent signals and increases the output to the brain. This heightened sensitivity of spinal neurons is called “central sensitization” [15]. Sensitization of spinal cord nociceptive neurons contributes to hypersensitive pain behaviors such as allodynia (pain produced by normally innocuous stimuli), hyperalgesia (heightened response to noxious stimuli) and spontaneous pain [16]. However, no direct link between the experimental observation of spinal cord neuron hyperexcitability and

the underlying mechanism of chronic pathological pain has been demonstrated [17]. Increasing evidence in the last decade has strengthened that chronic pain is a neuro-immune disorder that is caused by complex interactions between neurons, activated glial cells and inflammatory immune cells in the peripheral and CNS [18, 19]. This hypothesis suggests that the restoration of the balance between pro- and anti-inflammatory mechanisms may be used as a novel therapeutic approach to disrupt the development of chronic pain. Recent findings that support a relevant regulatory role for the TGF- $\beta$  family of cytokines in acute physiological nociception, neuroprotection and anti-inflammation in models of chronic pathological pain are discussed in the following.

### The Influence of BAMBI Deletion in Normal Nociception and in Models of Inflammatory and Neuropathic Pain

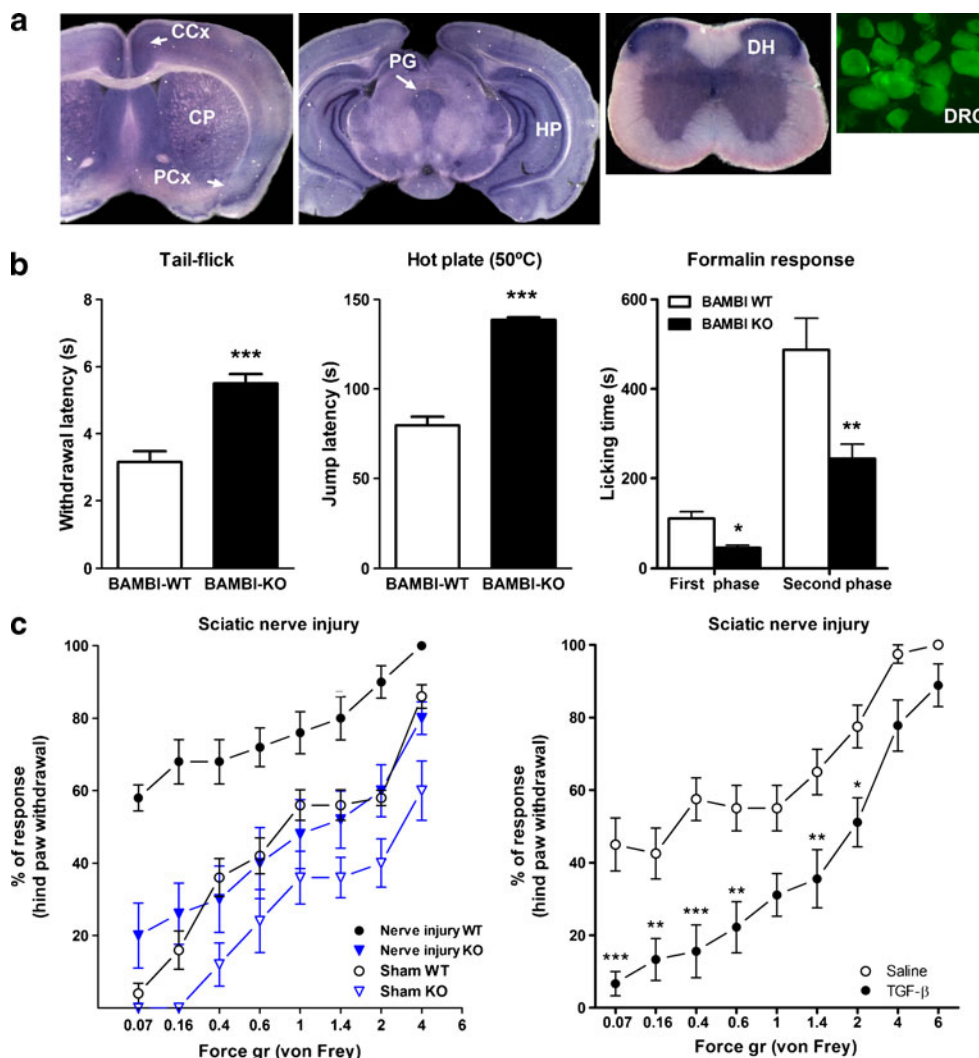
As mentioned previously, BAMBI is a kinase-deficient pseudoreceptor for TGF- $\beta$ s which prevents downstream signal transduction [10]. BAMBI, together with several type I TGF- $\beta$  receptors, is highly expressed in anti-nociception-relevant areas, such as the cingulate cortex, mesencephalic periaqueductal gray, spinal cord dorsal horn and DRG (Fig. 2) [20].

The induction of a gain in TGF- $\beta$  signaling through the deletion of the BAMBI gene has been a valuable strategy for unraveling the involvement of the TGF- $\beta$  family in pain control [20]. BAMBI-KO mice display attenuated nocifensive responses in acute pain models, regardless of the modality of the noxious stimuli (e.g. thermal, mechanical and chemical/inflammatory). Moreover, BAMBI-KO mice develop less mechanical allodynia in models of chronic neuropathic pain (Fig. 2). The hypoalgesic phenotype of BAMBI-KO mice can be reversed by the opioid antagonist naltrexone, which is indicative of an enhanced activity of the endogenous opioid system. Spinal cords from BAMBI-KO mice display higher expression levels of endogenous opioid peptide precursors, including  $\beta$ -endorphins (proopiomelanocortin: POMC) and enkephalins (proenkephalin: PENK), compared to wild-type mice. Opioid peptide precursors are under the transcriptional control of TGF- $\beta$  family members in cultured cells [21, 22]. Furthermore, exogenous TGF- $\beta$ s induce the expression of *POMC* and *PENK* in cultured spinal cord explants [20]. Therefore, an increase in TGF- $\beta$  signaling activity may lead to an increase in the transcription, expression and synaptic release of endogenous opioids and a hypoalgesic phenotype.

### The TGF- $\beta$ Subfamily Protects Against Nerve-Injury-Induced Neuropathic Pain

Pharmacological approaches have contributed to knowing the specific contribution of individual TGF- $\beta$  family members to





**Fig. 2** BAMBI expression in the mouse CNS and the hypoalgesic phenotype of BAMBI-KO mice. **a** Localization of BAMBI mRNA in brain and spinal cord sections (in situ hybridization using digoxigenin-labeled riboprobes) and BAMBI protein in DRG neurons (immunofluorescence). CCx cingulate cortex, PCx pyriform cortex, CP caudate-putamen nucleus, HP hippocampus, PG mesencephalic periaqueductal gray, DH dorsal horn of the spinal cord, DRG dorsal root ganglion. **b** Responses of BAMBI-KO and WT mice to acute painful stimuli. Two models of thermal nociception were used: The tail-flick test that examines spinal-mediated responses, and the hotplate test that examines both spinal and supraspinal-mediated responses. Chemical/inflammatory pain was induced by 20- $\mu$ l intraplantar injection of a 2% formalin solution in the left hind paw, and the cumulative time spent licking the paw was recorded within the first 5 min (first phase) and from 20 to 50 min after injection (second

phase). Data are means  $\pm$  SEM. \* $p$ <0.05, \*\* $p$ <0.01 and \*\*\* $p$ <0.001 versus BAMBI-WT mice (two-tailed Student's *t* test). **c** Development of neuropathic pain in response to crush injury of sciatic nerve. The left panel displays the mechanical allodynia evaluated with von Frey monofilaments on day 14 after nerve injury. Values are mean  $\pm$  SEM percentage of hind paw withdrawals elicited by mechanical stimuli of increasing strength in wild-type and BAMBI-KO mice subjected to sham operation or sciatic nerve injury. BAMBI-KO mice, compared with their wild-type littermates, are less sensitive to mechanical stimuli under basal conditions and develop attenuated allodynia following sciatic nerve injury. The right panel shows the anti-allodynic effect induced by a 2-week subcutaneous infusion of recombinant TGF- $\beta$ 1 in mice subjected to sciatic nerve injury, compared with saline. \* $p$ <0.05, \*\* $p$ <0.01 and \*\*\* $p$ <0.001 versus saline (Bonferroni test)

the control of nociceptive transmission. Echeverry et al. [23] demonstrated the protective role of TGF- $\beta$ 1 against nerve-injury-induced neuropathic pain. Thus, sustained intrathecal infusion of recombinant TGF- $\beta$ 1 during partial ligation of the sciatic nerve in rats significantly attenuates the development of mechanical allodynia and thermal hyperalgesia for 14 days. More importantly, from a therapeutic point of view,

TGF- $\beta$ 1 also produces a significant reduction in previously established hyperalgesia. Because TGF- $\beta$  can cross the blood–brain barrier (BBB) [24], we analyzed its effectiveness after systemic administration. As shown in Fig. 2, 2-week subcutaneous infusion of recombinant TGF- $\beta$ 1 significantly attenuates the development of mechanical allodynia in mice.

Echeverry et al. [23] attributed the biological effects of TGF- $\beta$ 1 to a milder neuroinflammatory response to injury in the spinal cord. These authors provided evidence that TGF- $\beta$ 1 exerted neuroprotective effects, inhibited the activation of spinal microglia and astrocytes and decreased the upregulation of pro-inflammatory cytokines, such as IL-1 $\beta$  and IL-6, within the spinal cord. These results are consistent with the anti-inflammatory properties of this cytokine [25, 26].

The contribution of peripheral mechanisms to the effects of TGF- $\beta$  is also important because several TGF- $\beta$  family members (mainly activin and BMPs) exert pro-nociceptive effects on peripheral nociceptors. Our results indicate that 2 weeks of systemic administration of a TGF- $\beta$  neutralizing antibody, which does not cross the BBB, significantly enhanced the development of mechanical allodynia in mice after sciatic nerve injury (our unpublished observations). These findings support the idea that TGF- $\beta$  also exerts peripheral anti-nociceptive effects and helps prevent peripheral nociceptor hypersensitization following nerve injury.

The capability of TGF- $\beta$  to maintain the integrity of the BBB has also been suggested as a protective mechanism against the development of pathological pain following peripheral inflammatory [27, 28] or neural injuries [29]. Systemic or intrathecal treatment with recombinant TGF- $\beta$ 1 prevents carrageenan- and nerve-injury-induced changes in endothelial tight junctions, as well as the functional disruption of the BBB. As a consequence, the extravasation of proteins, cytokines and a variety of inflammatory mediators and the infiltration of peripheral inflammatory cells in the spinal cord is significantly reduced. The mechanism involves ALK 5 receptors and the canonical Smad2/3 signaling pathway [27, 29].

In summary, the TGF- $\beta$  subfamily protects against pathological pain development by pleiotropic mechanisms. TGF- $\beta$  appears to promote the expression of endogenous opioids and inhibit the neuroimmune responses of glial cells and neurons in the spinal cord following peripheral injuries. In addition, undetermined peripheral mechanisms also contribute to its anti-allodynic effect. Even at this early stage of investigation, the evidence suggests that the modulation of TGF- $\beta$  signaling could be used as a novel pharmacological strategy for the control and treatment of chronic pain.

#### Neuropathic Pain is Dissimilarly Modulated by Individual Members of the BMP Subfamily

The role of the BMP family in the processing of nociceptive inputs within the CNS has been scarcely addressed, and different results were obtained depending on the specific BMP analyzed. Recent studies indicate that the use of recombinant BMP2 as bone inductor during

surgical spinal fusion elicits a profound Smad-mediated signaling response in the spinal cord and local DRG [30]. The direct access of BMP2 to the nervous system triggers a neuroinflammatory response that worsens the neurologic recovery and produces postoperative allodynia in rats [31]. This pro-algesic effect could have a clinical correlate as patients that received recombinant BMP2 were more likely to report postoperative radicular pain compared with patients whose fusion surgery did not include use of the protein [32]. However, the presence of a dural tear that would favor BMP2 diffusion into the spinal cord parenchyma did not increase the risk of radiculitis development or impair the neurologic recovery in a series of patients [33].

BMP4 has been reported to exert an indirect protective effect against hyperalgesia development. Neuropathic pain is a common harmful side effect of neuroepithelial stem cell transplantation therapy for spinal cord injury [34, 35]. The pretreatment of glial precursor cells with BMP4 (GDAs<sup>BMP</sup>) prevents the development of thermal hyperalgesia and mechanical allodynia in rats. Such protection may be related with the absence of dorsal horn sprouting of CGRP-immunoreactive C-fiber in mice treated with GDAs<sup>BMP</sup> [36]. In addition, the activation of BMP signaling pathways in GDAs<sup>BMP</sup> promotes the expression of glutamate transporter 1 and AKAP12 [37], which are genes relevant to preventing inflammatory and neuropathic pain [38] and to maintaining the BBB integrity [39]. By contrast, intrathecal delivery of BMP4 cDNA incorporated into an adenovirus vector does not modify somatosensory perceptions in mice subjected to spinal cord injury, while it promotes sensory axon regeneration [40]. Similarly, BMP7 overexpression in the sciatic nerve, DRG and spinal cord neurons by adenoviral gene transfer improves the functional recovery after injury but does not affect the development of neuropathic pain in rats [41].

#### Activin and BMP Retrograde Signals Modulate the Expression of the Neuropeptide CGRP by Nociceptors and Contribute to Neuroinflammatory Pain

The terminal differentiation of many developing neurons occurs after the innervation of their target cells. This differentiation is triggered by extrinsic, target-derived retrograde signals that are transduced by presynaptic cognate receptors. Target tissues contain neurotrophic growth factors that support neuronal survival and differentiation factors that are critical for the maturation of axonal projections and the acquisition of neuronal traits, such as the expression of distinct combinations of neurotransmitters [42, 43]. Epidermal keratinocytes have a dynamic neurochemical organization that plays a direct role in the modulation of sensory ending functions [44]. Retrograde

communication between the skin and nociceptive neurons by members of the TGF- $\beta$  family regulates the phenotypic identity of nociceptors and the circuit assembly. This communication also contributes to the plasticity that effects dynamic changes in nociceptor sensitivity in physiological and pathological situations.

TGF- $\beta$  receptors are broadly expressed by embryonic and adult neurons in trigeminal dorsal root sensory ganglia, which indicates the potential TGF- $\beta$  sensitivity of these cells [45–48]. In addition, members of the TGF- $\beta$  family are present in major nociceptor targets such as the skin [46, 49, 50].

A well-documented consequence of target-derived TGF- $\beta$  signals is the specification of the neurotransmitter phenotype of neuropeptide-containing DRG neurons. Glutamate is the major neurotransmitter released by nociceptors, but many nociceptive neurons also contain neuropeptide co-transmitters that help to transmit and modulate nociceptive inputs. Calcitonin gene-related peptide (CGRP) and substance P (SP) are the main neuropeptides that are expressed by an important subset of nociceptors that innervate skin and viscera [51]. Embryonic nociceptors contain no detectable CGRP mRNA or protein until their axons contact target tissues and the peripheral connections are functional, which indicates that peptidergic phenotype specification in nociceptive neurons is dependent on retrograde signals from targets [52, 53].

Studies *in vitro* have shown that skin-cell-derived factors induce *de novo* CGRP expression in embryonic DRG neurons that are isolated before peripheral target contact, and this effect is antagonized by anti-activin A antibody or the activin and BMP inhibitory protein, follistatin [45, 54–56]. Furthermore, the addition of recombinant activin A, BMP2, BMP4, BMP6 or BMP7 to cultured embryonic neurons that were dissociated from DRG induces a strong increase in the expression of CGRP by the specific subset of unmyelinated C-fiber peptidergic neurons. This effect involves the canonical Smad pathway [45, 56]. In postnatal DRG cultures, recombinant activin increases CGRP expression [56–59], while TGF- $\beta$ 1 and TGF- $\beta$ 2 increase the levels SP [59]. The effect of these cytokines on neuropeptide expression is synergistic with NGF, which led the authors to hypothesize that a cooperative interaction between Smads and NGF-dependent transcription factors further increases neuropeptide transcription [59]. Overall, these data support a role for TGF- $\beta$  family members as target-derived differentiation factors that contribute to the specification of the functional identity of neuropeptide-containing neurons during development. Moreover, sensory neurons remain sensitive to these cytokines during adulthood [46, 54, 58, 59].

The contribution of TGF- $\beta$  retrograde signals to the modulation of CGRP transcription by nociceptors *in vivo* is

highly relevant. CGRP conveys pain information from nociceptors to second-order neurons in the spinal cord, and it is also an important mediator of normal inflammatory pain response to injuries [60, 61]. CGRP is also involved in the development of hyperalgesia in inflammation- or nerve-injury-induced chronic pain [62]. Studies *in vivo* suggest that the conditional overexpression of BMP4 in mouse skin keratinocytes reduces the number of non-peptidergic neurons in sensory ganglia and the density of target innervation, but the nerve endings from CGRP peptidergic neurons were markedly increased [63]. This phenotype is consistent with the aforementioned effect of BMP signaling on CGRP expression *in vitro* [45]. The phenotypic changes that are induced by BMP4 overexpression are more prominent with increasing age, which suggests that the same retrograde communication between skin cells and sensory nerve endings is maintained in adults [62].

Activin is involved in the inflammatory response that is associated with the healing and repair of injured tissues in experimental adult animal models [56, 64–67]. Although the basal level of activin A in the skin is notably low, its expression is strongly increased following cutaneous excisional wound injury or under inflammation produced by complete Freund's adjuvant [56, 68, 69]. Under these experimental conditions, the CGRP-containing sensory neurons that innervate the skin wound region receive an important trophic influence from activin released by keratinocytes and inflammatory cells, which strongly enhances the expression of CGRP and other neuropeptides [56]. Furthermore, activin injection in the skin of adult naïve rats produces a tactile allodynia to mechanical stimulation that is associated with an increase in the proportion of CGRP-immunoreactive DRG neurons innervating the area of injection [57]. In addition,  $\beta$ -CGRP is released by skin keratinocytes, and its expression is regulated by autocrine/paracrine BMP signaling [62]. Importantly,  $\beta$ -CGRP is markedly increased in the skin of patients and mice with chronic inflammatory or neuropathic pain, which suggests an additional contribution of keratinocyte-dependent release of  $\beta$ -CGRP in regulating nociceptor plasticity under conditions of pathological pain [62].

The CGRP released by peripheral nerve endings and keratinocytes causes neurogenic vasodilatation, extravasation of serum factors important for wound healing, mast cell degranulation and platelet activation. The resulting accumulation of pro-inflammatory factors in the injured area plays an important role in the development and maintenance of peripheral sensitization of nociceptors and the phenotypic changes of DRG neurons that augment central transmission and peripheral sensitization. In addition, CGRP released by primary afferents contributes to the central sensitization of second-order neurons [70, 71]. The sensitization of the nociceptive system contributes to pain

hypersensitivity at the site of tissue damage and inflammation. Therefore, the modulation of CGRP-containing nociceptors by retrograde activin signaling promotes the heightened nocifensive behaviors that protect against further damage during the normal tissue healing process. An understanding of the role of activin and other TGF- $\beta$  family members in the CGRP-dependent sensitization of nociceptive signaling could provide further insights into the pathophysiology of highly prevalent pain diseases that involve this neuropeptide, such as migraine and other primary headaches [72].

#### Activin Sensitizes TRPV1 Ionic Currents in DRG Neurons and Produces Thermal Hyperalgesia in Mice

Transient receptor potential vanilloid 1 (TRPV1) is a non-selective cation channel that is expressed by polymodal nociceptors from somatic and visceral afferents, which are gated by noxious heat and irritant vanilloids, such as capsaicin and extracellular protons, and these nociceptors play a critical role in the development of heat hyperalgesia after inflammation [73] and visceral hyperalgesia in inflammatory bowel disease and rectal hypersensitivity [74]. Electrophysiological recordings of cultured DRG neurons suggest that recombinant activin A acutely potentiates capsaicin-induced ionic currents through TRPV1 by a mechanism that involves ALK4 receptors [48]. The epidermal injection of activin induces a rapid and short-lasting thermal hyperalgesia in wild-type mice, but this response is absent in TRPV1 null-mutant mice. Therefore, the positive modulation of TRPV1 by activin may be a novel mechanism underlying sustained inflammatory pain. Functionally, TRPV1 activation depends on a complicated balance of phosphorylation and dephosphorylation signals. Pro-nociceptive inflammatory mediators increase TRPV1 phosphorylation by the activation of multiple kinases, which sensitizes the TRPV1 response to noxious heat and regulates inflammatory hyperalgesia [75]. The non-canonical activation of PKC $\epsilon$  probably mediates acute TRPV1 sensitization by activin. In cultured DRG neurons, activin induces the translocation of PKC $\epsilon$  from the cytosol to the plasma membrane, and this effect is prevented by the blockade of ALK4 receptors. Moreover, a PKC $\epsilon$  translocation inhibitor peptide prevents activin-A-induced TRPV1 sensitization. Other protein kinases, such as PKA, MAPK, PI3K, c-src and other PKC isoforms, are not likely to play a role in TRPV1 sensitization because selective inhibitors of these kinases are ineffective [48].

From a therapeutic point of view, pharmacological approaches for the modulation of TRPV1 receptor function offers a promising means of pain relief at the nociceptor level. TRPV1 agonists produce a long-lasting desensitization of peripheral nerve terminals to noxious stimuli (nociceptor “defunctionalization”), which is the mechanism

that underlies the proven efficacy of TRPV1 agonists, such as capsaicin for neuropathic pain relief. TRPV1 antagonists also alleviate hyperalgesia associated with inflammatory pain [76]. Because activin has a sensitizing effect on TRPV1 channels, it would be useful to assess whether the inhibition of peripheral activin signaling interferes with the development of hyperalgesia under painful pathological conditions by reducing TRPV1 currents and whether exogenous activin induces nociceptor defunctionalization and long-lasting anti-nociception.

#### Conclusions and Perspectives

Members of the TGF- $\beta$  family play crucial roles in the pathophysiology of nociception, both at the peripheral sensory neurons and in the CNS. In models of inflammatory and neuropathic pain, the TGF- $\beta$  subfamily provides protection against the neuroinflammatory responses, prevents the disruption of the BBB integrity and favors the release of opioid analgesic mediators. The effects of the BMP family in the central processing of nociceptive inputs are largely unknown, and different results have been reported depending on the specific BMP analyzed. Recombinant BMP2 triggers a neuroinflammatory response that produces allodynia. Conversely, BMP4 could exert an indirect protective effect against hyperalgesia development. In the peripheral nervous system, activin and, presumably, BMPs promote heightened nocifensive behaviors that protect against further damage during the process of tissue healing. Activin also has a sensitizing effect on TRPV1 channels in DRG neurons, which elicits thermal hyperalgesia.

Further work is needed to identify the most promising components of the TGF- $\beta$  signaling pathways which might serve as viable therapeutic targets for the treatment of pathological pain. Also, due to the pleiotropic effects of these cytokines, a detailed investigation of the off-target effects that can potentially lead to unwanted toxicities is necessary before successful development of TGF- $\beta$ -based therapeutic strategies.

**Acknowledgments** This work was supported by grants from Instituto de Salud Carlos III (RD06/001/1016), Ministerio de Ciencia e Innovación (SAF2010-16894) and Fundación La Marató de TV3 (Grant 072131).

**Disclosures** None.

#### REFERENCES

1. Elliott AM, Smith BH, Penny KI, Smith WC, Chambers WA (1999) The epidemiology of chronic pain in the community. *Lancet* 354:1248–1252. doi:10.1016/S0140-6736(99)03057-3



2. Breivik H, Collett B, Ventafridda V, Cohen R, Gallacher D (2006) Survey of chronic pain in Europe: prevalence, impact on daily life, and treatment. *Eur J Pain* 10:287–333. doi:10.1016/j.ejpain.2005.06.009
3. Wharton K, Derynck R (2009) TGFbeta family signaling: novel insights in development and disease. *Development* 136:3691–3697. doi:10.1242/dev.040584
4. Schmierer B, Hill CS (2007) TGFβ-Smad signal transduction: molecular specificity and functional flexibility. *Nat Rev Mol Cell Biol* 8:970–982. doi:10.1038/nrm2297
5. Miyazono K, Maeda S, Imamura T (2005) BMP receptor signaling: transcriptional targets, regulation of signals, and signaling cross-talk. *Cytokine Growth Factor Rev* 16:251–263. doi:10.1016/j.cytogfr.2005.01.009
6. Zhang YE (2009) Non-Smad pathways in TGF-beta signaling. *Cell Res* 19:128–139. doi:10.1038/cr.2008.328
7. Kang JS, Liu C, Derynck R (2009) New regulatory mechanisms of TGF-beta receptor function. *Trends Cell Biol* 19:385–394. doi:10.1016/j.tcb.2009.05.008
8. Moustakas A, Heldin CH (2009) The regulation of TGFbeta signal transduction. *Development* 136(22):3699–3714. doi:10.1242/dev.030338
9. Umulis D, O'Connor MB, Blair SS (2009) The extracellular regulation of bone morphogenetic protein signaling. *Development* 136:3715–3728. doi:10.1242/dev.031534
10. Onichtchouk D, Chen YG, Dosch R, Gavantka V, Delius H, Massagué J, Niehrs C (1999) Silencing of TGF-beta signalling by the pseudoreceptor BAMBI. *Nature* 401:480–485. doi:10.1038/46794
11. Itoh S, ten Dijke P (2007) Negative regulation of TGF-β receptor/Smad signal transduction. *Curr Opin Cell Biol* 19:176–184. doi:10.1016/j.ccb.2007.02.015
12. Basbaum AI, Bautista DM, Scherrer G, Julius D (2009) Cellular and molecular mechanisms of pain. *Cell* 139:267–284. doi:10.1016/j.cell.2009.09.028
13. Cervero F (2009) Pain: friend or foe? A neurobiologic perspective: the 2008 Bonica Award Lecture. *Reg Anesth Pain Med* 34:569–574. doi:10.1097/AAP.0b013e3181b4c517
14. Bouhassira D, Lantéri-Minet M, Attal N, Laurent B, Touboul C (2008) Prevalence of chronic pain with neuropathic characteristics in the general population. *Pain* 136:380–387. doi:10.1016/j.pain.2007.08.013
15. D'Mello R, Dickenson AH (2008) Spinal cord mechanisms of pain. *Br J Anaesth* 101:8–16. doi:10.1093/bja/aen088
16. Costigan M, Scholz J, Woolf CJ (2009) Neuropathic pain: a maladaptive response of the nervous system to damage. *Annu Rev Neurosci* 32:1–32. doi:10.1146/annurev.neuro.051508.135531
17. Cervero F (2009) Spinal cord hyperexcitability and its role in pain and hyperalgesia. *Exp Brain Res* 196:129–137. doi:10.1007/s00221-009-1789-2
18. Milligan ED, Watkins LR (2009) Pathological and protective roles of glia in chronic pain. *Nat Rev Neurosci* 10:23–36. doi:10.1038/nrn2533
19. Austin PJ, Moalem-Taylor G (2010) The neuro-immune balance in neuropathic pain: involvement of inflammatory immune cells, immune-like glial cells and cytokines. *J Neuroimmunol* 229:26–50. doi:10.1016/j.jneuroim.2010.08.013
20. Tramullas M, Lantero A, Díaz A, Morchón N, Merino D, Villar A, Buscher D, Merino R, Hurlé JM, Izpisua-Belmonte JC, Hurlé MA (2010) BAMBI (bone morphogenetic protein and activin membrane-bound inhibitor) reveals the involvement of the transforming growth factor-beta family in pain modulation. *J Neurosci* 30:1502–1511. doi:10.1523/JNEUROSCI.2584-09.2010
21. Kamphuis S, Kavelaars A, Brooimans R, Kuis W, Zegers BJ, Heijnen CJ (1997) T helper 2 cytokines induce preproenkephalin mRNA expression and proenkephalin A in human peripheral blood mononuclear cells. *J Neuroimmunol* 79:91–99. doi:10.1016/S0165-5728(97)00113-6
22. Nudi M, Ouimette JF, Drouin J (2005) Bone morphogenic protein (Smad)-mediated repression of proopiomelanocortin transcription by interference with Pitx/Tpit activity. *Mol Endocrinol* 19:1329–1342. doi:10.1210/me.2004-0425
23. Echeverry S, Shi XQ, Haw A, Liu H, Zhang ZW, Zhang J (2009) Transforming growth factor-beta1 impairs neuropathic pain through pleiotropic effects. *Mol Pain* 5:16. doi:10.1186/1744-8069-5-16
24. McLennan IS, Weible MW 2nd, Hendry IA, Koishi K (2005) Transport of transforming growth factor-beta 2 across the blood-brain barrier. *Neuropharmacology* 48:274–282. doi:10.1016/j.neuropharm.2004.10.005
25. Bottner M, Kriegstein K, Unsicker K (2000) The TGF-βs: structure, signalling and roles in nervous system development and functions. *J Neurochem* 75:2227–2240. doi:10.1046/j.1471-4159.2000.0752227.x
26. Brionne TC, Tesseur I, Masliah E, Wyss-Coray T (2003) Loss of TGF-beta 1 leads to increased neuronal cell death and microglialosis in mouse brain. *Neuron* 40:1133–1145. doi:10.1016/S0896-6273(03)00766-9
27. Ronaldson PT, Demarco KM, Sanchez-Covarrubias L, Solinsky CM, Davis TP (2009) Transforming growth factor-beta signaling alters substrate permeability and tight junction protein expression at the blood-brain barrier during inflammatory pain. *J Cereb Blood Flow Metab* 29:1084–1098. doi:10.1038/jcbfm.2009.32
28. Ronaldson PT, Finch JD, Demarco KM, Quigley CE, Davis TP (2011) Inflammatory pain signals an increase in functional expression of organic anion transporting polypeptide 1a4 at the blood-brain barrier. *J Pharmacol Exp Ther* 336:827–839. doi:10.1124/jpet.110.174151
29. Echeverry S, Shi XQ, Rivest S, Zhang J (2011) Peripheral nerve injury alters blood-spinal cord barrier functional and molecular integrity through a selective inflammatory pathway. *J Neurosci* 31:10819–11028. doi:10.1523/JNEUROSCI.1642-11.2011
30. Dmitriev AE, Farhang S, Lehman RA Jr, Ling GS, Symes AJ (2010) Bone morphogenetic protein-2 used in spinal fusion with spinal cord injury penetrates intrathecally and elicits a functional signaling cascade. *Spine J* 10:16–25. doi:10.1016/j.spinee.2009.10.003
31. Dmitriev AE, Lehman RA Jr, Symes AJ (2011) Bone morphogenetic protein-2 and spinal arthrodesis: the basic science perspective on protein interaction with the nervous system. *Spine J* 11:500–505. doi:10.1016/j.spinee.2011.05.002
32. Carragee EJ, Hurwitz EL, Weiner BK (2011) A critical review of recombinant human bone morphogenetic protein-2 trials in spinal surgery: emerging safety concerns and lessons learned. *Spine J* 11:471–491. doi:10.1016/j.spinee.2011.04.023
33. Glassman SD, Gum JL, Crawford CH 3rd, Shields CB, Carreon LY (2011) Complications with recombinant human bone morphogenetic protein-2 in posterolateral spine fusion associated with a dural tear. *Spine J* 11:522–526. doi:10.1016/j.spinee.2010.05.016
34. Hofstetter CP, Holmström NA, Lilja JA, Schweinhardt P, Hao J, Spenger C, Wiesenfeld-Hallin Z, Kurpad SN, Frisén J, Olson L (2005) Allodynia limits the usefulness of intraspinal neural stem cell grafts; directed differentiation improves outcome. *Nat Neurosci* 8:346–353. doi:10.1038/nn1405
35. Macias MY, Syring MB, Pizzi MA, Crowe MJ, Alexanian AR, Kurpad SN (2006) Pain with no gain: allodynia following neural stem cell transplantation in spinal cord injury. *Exp Neurol* 201:335–348. doi:10.1016/j.expneurol.2006.04.035
36. Davies JE, Pröschel C, Zhang N, Noble M, Mayer-Pröschel M, Davies SJ (2008) Transplanted astrocytes derived from BMP- or CNTF-treated glial-restricted precursors have opposite effects on recovery and allodynia after spinal cord injury. *J Biol* 7:24. doi:10.1186/jbiol85

37. Davies SJ, Shih CH, Noble M, Mayer-Proschel M, Davies JE, Proschel C (2011) Transplantation of specific human astrocytes promotes functional recovery after spinal cord injury. *PLoS One* 6:e17328. doi:10.1371/journal.pone.0017328
38. Maeda S, Kawamoto A, Yatani Y, Shirakawa H, Nakagawa T, Kaneko S (2008) Gene transfer of GLT-1, a glial glutamate transporter, into the spinal cord by recombinant adenovirus attenuates inflammatory and neuropathic pain in rats. *Mol Pain* 4:65. doi:10.1186/1744-8069-4-65
39. Choi YK, Kim JH, Kim WJ, Lee HY, Park JA, Lee SW, Yoon DK, Kim HH, Chung H, Yu YS (2007) Kim KW (2007) AKAP12 regulates human blood-retinal barrier formation by downregulation of hypoxia-inducible factor-1alpha. *J Neurosci* 27:4472–4481. doi:10.1523/JNEUROSCI.5368-06.2007
40. Parikh P, Hao Y, Hosseinkhani M, Patil SB, Huntley GW, Tessier-Lavigne M, Zou H (2011) Regeneration of axons in injured spinal cord by activation of bone morphogenetic protein/Smad1 signaling pathway in adult neurons. *Proc Natl Acad Sci U S A* 108:E99–E107. doi:10.1073/pnas.1100426108
41. Tsai MJ, Pan HA, Liou DY, Weng CF, Hoffer BJ, Cheng H (2010) Adenoviral gene transfer of bone morphogenetic protein-7 enhances functional recovery after sciatic nerve injury in rats. *Gene Ther* 17:1214–1224. doi:10.1038/gt.2010.72
42. Hippenmeyer S, Kramer I, Arber S (2004) Control of neuronal phenotype: what targets tell the cell bodies. *Trends Neurosci* 27:482–488. doi:10.1016/j.tins.2004.05.012
43. Sanyal S, Kim SM, Ramaswami M (2004) Retrograde regulation in the CNS; neuron-specific interpretations of TGF-beta signaling. *Neuron* 41:845–848. doi:10.1016/S0896-6273(04)00152-7
44. Lumpkin EA, Caterina MJ (2007) Mechanisms of sensory transduction in the skin. *Nature* 445:858–865. doi:10.1038/nature05662
45. Ai X, Cappuzzello J, Hall AK (1999) Activin and bone morphogenetic proteins induce calcitonin gene-related peptide in embryonic sensory neurons in vitro. *Mol Cell Neurosci* 14:506–518. doi:10.1006/mcne.1999.0798
46. Hall AK, Burke RM, Anand M, Dinsio KJ (2002) Activin and bone morphogenetic proteins are present in perinatal sensory neuron target tissues that induce neuropeptides. *J Neurobiol* 52:52–60. doi:10.1002/neu.10068
47. Zhang D, Mehler MF, Song Q, Kessler JA (1998) Development of bone morphogenetic protein receptors in the nervous system and possible roles in regulating trkC expression. *J Neurosci* 18:3314–3326. doi:10.1002/neu.10068
48. Zhu W, Xu P, Cuascut FX, Hall AK, Oxford GS (2007) Activin acutely sensitizes dorsal root ganglion neurons and induces hyperalgesia via PKC-mediated potentiation of transient receptor potential vanilloid 1. *J Neurosci* 27:13770–13780. doi:10.1523/JNEUROSCI.3822-07.2007
49. Winnier G, Blessing M, Labosky PA, Hogan BL (1995) Bone morphogenetic protein-4 is required for mesoderm formation and patterning in the mouse. *Genes Dev* 9:2105–2116. doi:10.1101/gad.9.17.2105
50. Takahashi H, Ikeda T (1996) Transcripts for two members of the transforming growth factor-beta superfamily BMP-3 and BMP-7 are expressed in developing rat embryos. *Dev Dyn* 207:439–449. doi:10.1002/(SICI)1097-0177(1996)2
51. Yu LC, Hou JF, Fu FH, Zhang YX (2009) Roles of calcitonin gene-related peptide and its receptors in pain-related behavioral responses in the central nervous system. *Neurosci Biobehav Rev* 33:1185–1191. doi:10.1016/j.neubiorev.2009.03.009
52. Hall AK, Ai X, Hickman GE, MacPhedran SE, Nduaguba CO, Robertson CP (1997) The generation of neuronal heterogeneity in a rat sensory ganglion. *J Neurosci* 17:2775–2784
53. Marti E, Gibson SJ, Polak JM, Facer P, Springall DR, Van Aswegen G, Aitchison M, Koltzenburg M (1987) Ontogeny of peptide- and amine-containing neurones in motor, sensory, and autonomic regions of rat and human spinal cord, dorsal root ganglia, and rat skin. *J Comp Neurol* 266:332–359
54. Hall AK, Dinsio KJ, Cappuzzello J (2001) Skin cell induction of calcitonin gene-related peptide in embryonic sensory neurons in vitro involves activin. *Dev Biol* 229:263–270. doi:10.1006/dbio.2000.9966
55. Hamza MA, Higgins DM, Ruyechan WT (2007) Two alphaherpesvirus latency-associated gene products influence calcitonin gene-related peptide levels in rat trigeminal neurons. *Neurobiol Dis* 25:553–560. doi:10.1016/j.nbd.2006.10.016
56. Cruise BA, Xu P, Hall AK (2004) Wounds increase activin in skin and a vasoactive neuropeptide in sensory ganglia. *Dev Biol* 271:1–10. doi:10.1016/j.ydbio.2004.04.003
57. Xu P, Van Slambrouck C, Berti-Mattera L, Hall AK (2005) Activin induces tactile allodynia and increases calcitonin gene-related peptide after peripheral inflammation. *J Neurosci* 25:9227–9235. doi:10.1523/JNEUROSCI.3051-05.2005
58. Xu P, Hall AK (2006) The role of activin in neuropeptide induction and pain sensation. *Dev Biol* 299:303–309. doi:10.1016/j.ydbio.2006.08.026
59. Xu P, Hall AK (2007) Activin acts with nerve growth factor to regulate calcitonin gene-related peptide mRNA in sensory neurons. *Neuroscience* 150:665–674. doi:10.1016/j.neuroscience.2007.09.041
60. Salmon AM, Damaj MI, Marubio LM, Epping-Jordan MP, Merlo-Pich E, Changeux JP (2001) Altered neuroadaptation in opiate dependence and neurogenic inflammatory nociception in alpha CGRP-deficient mice. *Nat Neurosci* 4:357–358. doi:10.1038/86001
61. Zhang Z, Winborn CS, Marquez de Prado B, Russo AF (2007) Sensitization of calcitonin gene-related peptide receptors by receptor activity-modifying protein-1 in the trigeminal ganglion. *J Neurosci* 27:2693–2703. doi:10.1523/JNEUROSCI.4542-06.2007
62. Hou Q, Barr T, Gee L, Vickers J, Wymer J, Borsani E, Rodella L, Getsios S, Burdo T, Eisenberg E, Guha U, Lavker R, Kessler J, Chittur S, Fiorino D, Rice F, Albrecht P. Keratinocyte expression of calcitonin gene-related peptide  $\beta$ : implications for neuropathic and inflammatory pain mechanisms. *Pain* 152:2036–2051. doi:10.1016/j.pain.2011.04.033
63. Guha U, Gomes WA, Samanta J, Gupta M, Rice FL, Kessler JA (2004) Target-derived BMP signaling limits sensory neuron number and the extent of peripheral innervation in vivo. *Development* 131:1175–1186. doi:10.1242/dev.01013
64. Hübner G, Hu Q, Smola H, Werner S (1996) Strong induction of activin expression after injury suggests an important role of activin in wound repair. *Dev Biol* 173:490–498. doi:10.1006/dbio.1996.0042
65. Wankell M, Munz B, Hübner G, Hans W, Wolf E, Goppelt A, Werner S (2001) Impaired wound healing in transgenic mice overexpressing the activin antagonist follistatin in the epidermis. *EMBO J* 20:5361–5372. doi:10.1093/emboj/20.19.5361
66. Bamberger C, Schärer A, Antsiferova M, Tytsen B, Pankow S, Müller M, Rüllicke T, Paus R, Werner S (2005) Activin controls skin morphogenesis and wound repair predominantly via stromal cells and in a concentration-dependent manner via keratinocytes. *Am J Pathol* 167:733–747. doi:10.1016/S0002-9440(10)62047-0
67. Antsiferova M, Klatte JE, Bodó E, Paus R, Jorcano JL, Matzuk MM, Werner S, Kögel H (2009) Keratinocyte-derived follistatin regulates epidermal homeostasis and wound repair. *Lab Invest* 89:131–141. doi:10.1038/labinvest.2008.120
68. Hübner G, Hu Q, Smola H (1996) Werner S (1996) Strong induction of activin expression after injury suggests an important role of activin in wound repair. *Dev Biol* 173(2):490–498. doi:10.1006/dbio.1996.0042
69. Sulyok S, Wankell M, Alzheimer C, Werner S (2004) Activin: an important regulator of wound repair, fibrosis, and neuroprotection. *Mol Cell Endocrinol* 225:127–132. doi:10.1016/j.mce.2004.07.011

70. Zhang L, Hoff AO, Wimalawansa SJ, Cote GJ, Gagel RF, Westlund KN (2001) Arthritic calcitonin/alpha calcitonin gene-related peptide knockout mice have reduced nociceptive hypersensitivity. *Pain* 89:265–273. doi:[10.1016/S0304-3959\(00\)00378-X](https://doi.org/10.1016/S0304-3959(00)00378-X)
71. Benemei S, Nicoletti P, Capone JG, Geppetti P (2009) CGRP receptors in the control of pain and inflammation. *Curr Opin Pharmacol* 9:9–14. doi:[10.1016/j.coph.2008.12.007](https://doi.org/10.1016/j.coph.2008.12.007)
72. Ho TW, Edvinsson L, Goadsby PJ (2010) CGRP and its receptors provide new insights into migraine pathophysiology. *Nat Rev Neurol* 6:573–582. doi:[10.1038/nrneurol.2010.127](https://doi.org/10.1038/nrneurol.2010.127)
73. Davis JB, Gray J, Gunthorpe MJ, Hatcher JP, Davey PT, Overend P, Harries MH, Latcham J, Clapham C, Atkinson K, Hughes SA, Rance K, Grau E, Harper AJ, Pugh PL, Rogers DC, Bingham S, Randall A, Sheardown SA (2000) Vanilloid receptor-1 is essential for inflammatory thermal hyperalgesia. *Nature* 405:183–187. doi:[10.1038/35012076](https://doi.org/10.1038/35012076)
74. Cervero F, Laird JMA (2004) Understanding the signaling and transmission of visceral nociceptive events. *J Neurobiol* 61:45–54. doi:[10.1002/neu.20084](https://doi.org/10.1002/neu.20084)
75. Ma W, Quirion R (2007) Inflammatory mediators modulating the transient receptor potential vanilloid 1 receptor: therapeutic targets to treat inflammatory and neuropathic pain. *Expert Opin Ther Targets* 11:307–320. doi:[10.1517/14728222.11.3.307](https://doi.org/10.1517/14728222.11.3.307)
76. Gunthorpe MJ, Chizh BA (2009) Clinical development of TRPV1 antagonists: targeting a pivotal point in the pain pathway. *Drug Discov Today* 14:56–67. doi:[10.1016/j.drudis.2008.11.005](https://doi.org/10.1016/j.drudis.2008.11.005)